



ALLIANCE™

<https://www.globalseafood.org>Health &
Welfare

Differing water salinities can shift bacterial composition in RAS shrimp production

17 January 2022

By Dr. Julia Bauer , Dr. Felix Teitge , Lisa Neffe, M.S. , Dr. Mikolaj Adamek , Dr. Arne Jung , Christina Pepler , Dr. Dieter Steinhagen and Dr. Verena Jung-Schroers

Test results show bacterial composition differed significantly depending on water salinity

The Pacific white shrimp (*Litopenaeus vannamei*) is the most cultivated shrimp species worldwide. Its native habitat is the eastern Pacific coast of South America with an average water temperature of above 20 degrees-C and a salinity of 34–37 ppt. In aquaculture, this species is typically produced in ponds at water temperatures between 26–32 degrees-C, mostly in South America and Asia. By developing land-based recirculating aquaculture systems (RAS), the production of *L. vannamei* has become possible also in northern European countries and North America in inland farms independent from natural sea water.

The main challenges of keeping shrimp in RAS are the maintenance of high water temperatures and appropriate salinities. By using waste heat from biogas plants, the heating of RAS water can be achieved in a sustainable way. As *L. vannamei* are very tolerant against low or moderate salinity levels, RAS for on-growing shrimps after the post-larval stage are often operated at a salinity of 10–15 ppt to



This study evaluated the effect of different water salinities on *Vibrio* species composition in *L. vannamei* shrimp RAS production. Results showed that bacterial composition differed significantly depending on water salinity, with shifts in *Vibrio* populations towards pathogenic species with reduced salinities, and possibly increased risks for shrimp disease and potential food safety issues. Photo by Darryl Jory.

reduce costs for artificial sea salt and the amount of salty wastewater. This contributes further to the sustainability of a local shrimp production. However, even though shrimp tolerate brackish salinities, their innate immunity can be suppressed when the salinity in water is reduced, especially when the ion composition is suboptimal.

In marine and brackish water RAS, a great variety of *Vibrio* spp. can be found. These are ubiquitous bacteria in seawater and brackish water, with a share of the bacterial community of up to 40 percent. Several *Vibrio* spp. form a part of the natural microflora of fish and shellfish, but some species can also act as facultative (discretionary) pathogens for shrimp or finfish. In particular, *V. alginolyticus*, *V. campbellii*, *V. harveyi*, *V. owensii* and *V. parahaemolyticus* are potential pathogens for shrimp. Some species of *Vibrio* are known to be potentially pathogenic for humans as well, and especially *V. cholerae*, *V. parahaemolyticus* and *V. vulnificus* are major causes of seafood-borne illnesses.

This article – adapted and summarized from the **original publication** (<https://doi.org/10.1111/jfd.13270>). [Bauer, J. et al. 2021. Impact of a reduced water salinity on the composition of *Vibrio* spp. in recirculating aquaculture systems for Pacific white shrimp (*Litopenaeus vannamei*) and its possible risks for shrimp health and food safety. J. Fish Diseases 44(1):89-105] –

reports on an investigation of the influence of lowered salinity in shrimp RAS on the composition of *Vibrio* spp., comparing salinities of 15 and 30 ppt. Additionally, the pathogenicity factors of the main detected species at each salinity were analyzed to assess the potential risk for shrimp health and food safety.



(<https://bspcertification.org/>).

Study setup

Water samples analyzed originated from six different RAS stocked with *L. vannamei* of different ages and under different production conditions. Three RAS were maintained at a salinity of around 30 ppt, and three RAS were maintained at a salinity of around 15 ppt. RAS 1, 3, 5 and 6 were located in different research facilities. RAS 1 and RAS 3 consisted of three 100-liter holding tanks. RAS 5 and RAS 6 consisted of three 70-liter holding tanks.

All RAS in research facilities were stocked with specific pathogen-free *L. vannamei* from a commercial hatchery (Florida, USA). Samples from RAS 2 and RAS 4 originated from commercial production facilities for *L. vannamei* located in Northern Europe. RAS 2 consisted of two 7,000-liter holding tanks, and RAS 4 consisted of four 88,000-liter holding tanks. Water samples from each holding tank of both RAS were analyzed.

For detailed information on the sampling and analyses for total bacterial numbers and species identification; pathogenicity and other analyses; and statistical analysis, refer to the original publication.

Results and discussion

In recent years, the production of tropical shrimp like *L. vannamei* is growing (at relatively small scale) in Northern European countries in land-based recirculation systems far away from the sea. The maintenance of an optimal chemical and microbiological water quality is crucial for shrimp health in RAS, especially when only low amounts of water are exchanged during the production cycle. Issues in water temperature and chemical water quality, like increased ammonia and nitrite concentrations as well as suboptimal pH levels, might lead to a higher susceptibility of the animals to bacterial infections, for example, caused by *V. alginolyticus*.

Changes in water salinity are crucial for shrimp health as well. *L. vannamei* reportedly grows best at 24 ppt but they can tolerate a wide range of salinities, up to 45 ppt. This is exploited in RAS production, where the salinity is often reduced to 10–15 ppt after the post-larval stage to reduce the costs for artificial sea salt and also to lower the salt content in wastewater. When culturing *L. vannamei* at higher salinities from 25 up to 45 ppt, the animals maintain good growth, but keeping them at low salinities requires the maintenance of optimal potassium (K⁺) and magnesium (Mg²⁺) concentrations in the artificial sea water to achieve optimal weight gains. Furthermore, the ratios of Na: K (sodium: potassium) and Mg: Ca (magnesium: calcium) also have to be considered as possible disruptive factors and need to be kept within optimal ranges. If these ions are not adjusted in shrimp aquaculture, low salinity may lead to a higher susceptibility to bacterial pathogens like *V. alginolyticus*.

All isolates of *V. owensii* examined in the present study tested negative for the AHPND toxin genes (pirA,B) and only a few of the examined isolates tested positive for other pathogenicity factors. Nevertheless, as *V. owensii* might also carry the AHPND toxins and could only be detected in one of the RAS with a low water salinity of 15 ppt, a reduced salinity seems to be a risk factor for infections of shrimp by *V. owensii*.

All examined *V. campbellii* isolates in the present study tested negative for AHPND toxin genes and all other analyzed pathogenicity factors. *V. campbellii* was only detected in the RAS run at 15 ppt, so it could be a potential threat to shrimp in systems with lowered salinity.

Perspectives

Our results show that the *Vibrio* population in water of shrimp RAS differed significantly depending on water salinity. In RAS at a lower salinity of 15 ppt, higher numbers of the potential pathogenic species *V. parahaemolyticus*, *V. owensii* and *V. campbellii* were detected, all of which might produce the toxins that cause Acute Hepatopancreatic Necrosis Disease (AHPND) in shrimp. In water with a high salinity of 30 ppt, these species were virtually absent.

The toxins were not detected in any of the isolates, but as multiple other pathogenicity factors were present, especially in the examined *V. parahaemolyticus* isolates, it can be concluded that a reduced salinity may lead to a shift in *Vibrio* populations towards pathogenic species and thereby increase the risk of disease outbreaks in shrimp in RAS. When considering reducing the salinity in RAS for shrimp production, not only the negative effects on shrimp innate immunity but also the shift in *Vibrio* species composition should be considered.

Authors



DR. JULIA BAUER

Fish Disease Research Unit, University of Veterinary Medicine Hannover, Hannover, Germany



DR. FELIX TEITGE

Fish Disease Research Unit, University of Veterinary Medicine Hannover, Hannover, Germany



LISA NEFFE, M.S.

Fish Disease Research Unit, University of Veterinary Medicine Hannover, Hannover, Germany



DR. MIKOLAJ ADAMEK

Fish Disease Research Unit, University of Veterinary Medicine Hannover, Hannover, Germany



DR. ARNE JUNG

Clinic for Poultry, University of Veterinary Medicine Hannover, Hannover, Germany



CHRISTINA PEPLER

Polyplan-Kreikenbaum Group GmbH, Bremen, Germany



DR. DIETER STEINHAGEN

Fish Disease Research Unit, University of Veterinary Medicine Hannover, Hannover, Germany



DR. VERENA JUNG-SCHROERS

Corresponding author
Fish Disease Research Unit, University of Veterinary Medicine Hannover, Hannover, Germany

Verena.jung-schroers@tiho-hannover.de (<mailto:Verena.jung-schroers@tiho-hannover.de>)

Copyright © 2024 Global Seafood Alliance

All rights reserved.